

## Media for protozoa

### 1/2Waris H+Si

HEPES	11.915 mg
KNO <sub>3</sub>	5 mg
MgSO <sub>4</sub> · 7H <sub>2</sub> O	1 mg
(NH <sub>4</sub> ) <sub>2</sub> HPO <sub>4</sub>	1 mg
Ca(NO <sub>3</sub> ) <sub>2</sub> · 4H <sub>2</sub> O	5 mg
Waris P-2	0.05 mL
Waris EDTA	0.05 mL
Vitamin B <sub>12</sub>	1 µg
Biotin	0.05 µg
Thiamine HCl	5 µg
Niacinamide	0.005 µg
Soil extract	0.5 mL
Na <sub>2</sub> SiO <sub>3</sub> · 9H <sub>2</sub> O	2.842 mg
Distilled water	99.4 mL
PH 7.0	

Waris H is diluted half strength and added Na<sub>2</sub>SiO<sub>3</sub> · 9H<sub>2</sub>O.

#### Reference

McFadden, G. I., Melkonian, M. 1986 Use of HEPES buffer for microalgal culture media and fixation for electron microscopy. *Phycologia*, **25**, 551-557.

### Waris P-2

Na <sub>2</sub> EDTA · 2H <sub>2</sub> O	0.3 g
H <sub>3</sub> BO <sub>3</sub>	0.114 g
MnCl <sub>2</sub> · 4H <sub>2</sub> O	14.4 mg
ZnSO <sub>4</sub> · 7H <sub>2</sub> O	2.1 mg
CoCl <sub>2</sub> · 6H <sub>2</sub> O	0.4 mg
Distilled water	100 mL

#### Reference

McFadden, G. I., Melkonian, M. 1986 Use of HEPES buffer for microalgal culture media and fixation for electron microscopy. *Phycologia*, **25**, 551-557.

## Media for protozoa

### Waris EDTA

EDTA	0.522	g
FeSO <sub>4</sub> · 7H <sub>2</sub> O	0.498	g
KOH (1 mol/L solution)	5.4	mL
Distilled water	94.6	mL

EDTA and FeSO<sub>4</sub> · 7H<sub>2</sub>O is heated for 30 min (100°C); KOH is added to the cooled mixture.

### Reference

McFadden, G. I., Melkonian, M. 1986 Use of Hepes buffer for microalgal culture media and fixation for electron microscopy. *Phycologia*, **25**, 551-557.